

Confocal Microscopy – Olympus FV1000 Confocal Laser Scanning Microscope.

- Confocal Microscope set up
 - o Lasers
 - Argon-ion – multi line 458, 488, 515nm wavelength
 - Helium-neon – Green – 543nm, Red – 633nm
 - UV – 405nm
 - o Spectral Scan Unit
 - o Spectral detection system uses diffraction grating with variable slit – allows the separation of fluorochromes down to 2nm.
 - o Photomultiplier tube detector (PMT)
 - o Amplification of signals
 - o Pinhole – it's all about the pinhole

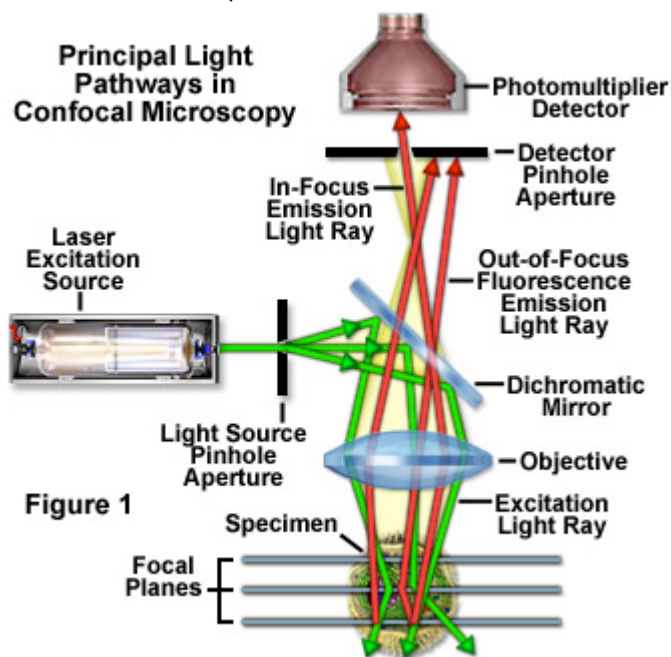
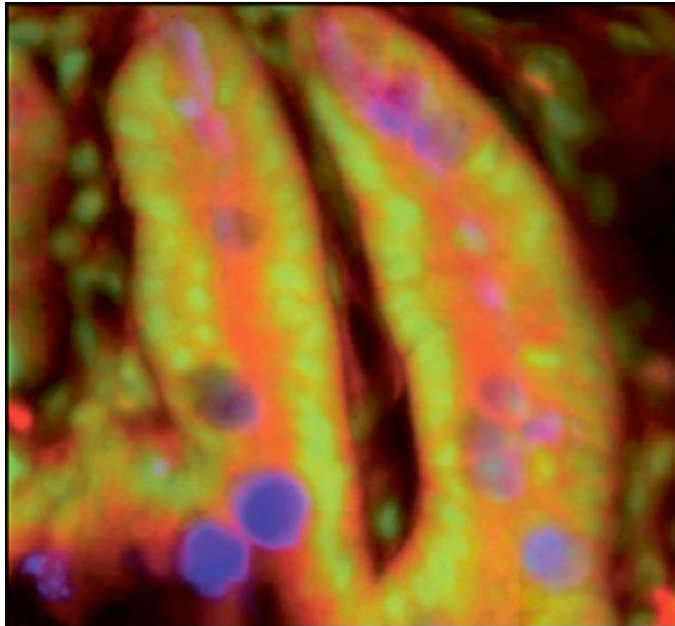
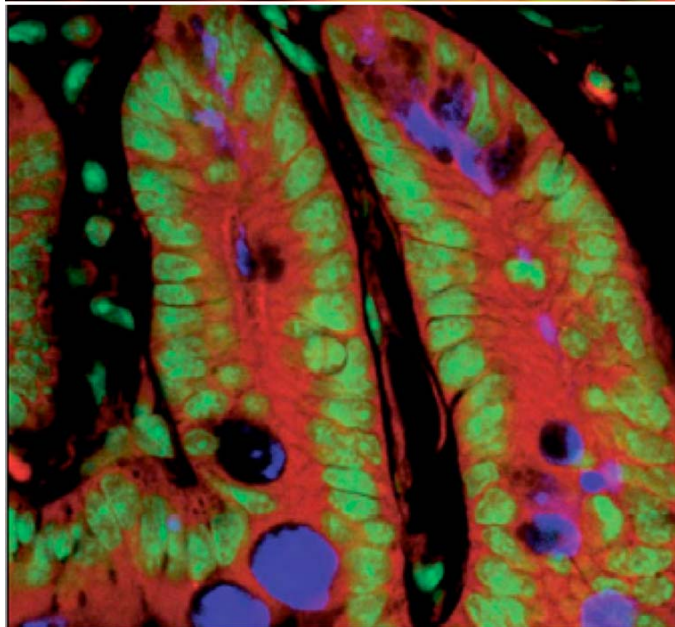


Figure 1

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- o (From <http://www.microscopyu.com/articles/confocal/confocalintrobasics.html>)



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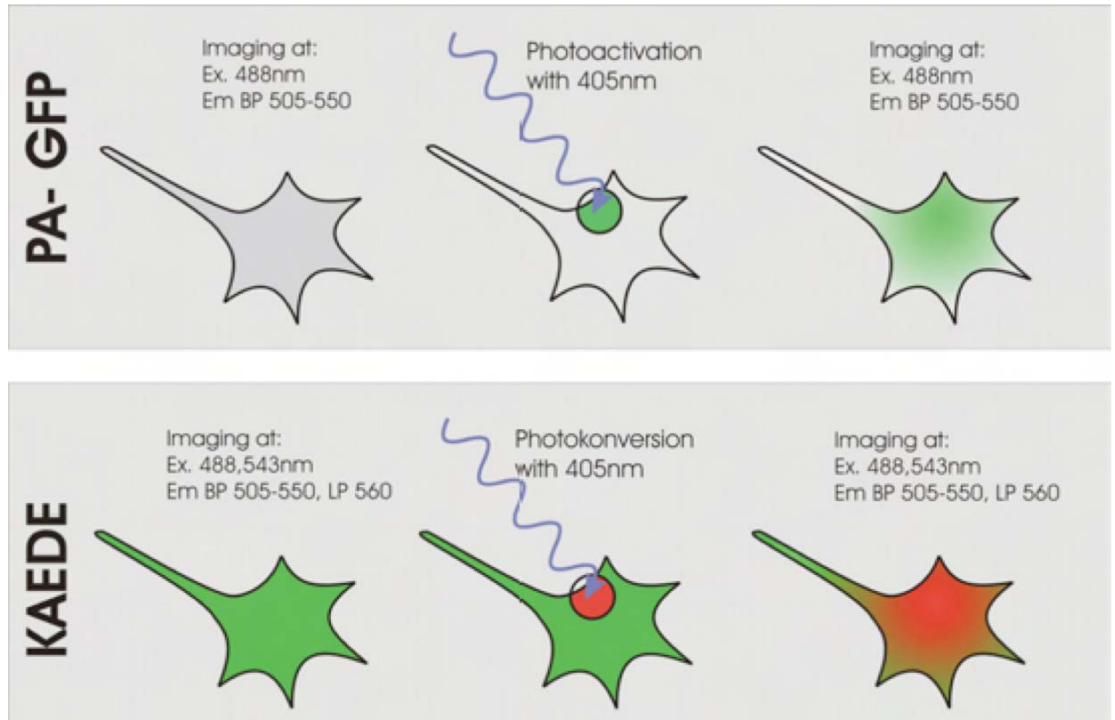
(from <http://www.dbs.nus.edu.sg/research/facilities/confocal/LSM%20510%20META%20training-DBS-070605.pdf>)

- Other parts
 - Microscope – upright/inverted
 - PC with a fair bit of memory
 - Optional extras – Live cell setup

- The unique scan head set up allows scanning:
 - XY
 - XYZ (3 dimensions)
 - XZ

- XYt (over time)
 - XYZt (over 3 dimensions and time!)
 - XYt λ (over time and different wavelengths!!)
- Multiple fluorescent probes
 - Molecular Probes control slide – Bovine pulmonary arterial endothelial cells: DAPI = nucleus, BODIPY = microtubules, TEXAS RED = actin
 - Labelled antibodies
 - GFPs – genetic code known so that genetically fused proteins are possible.
- Applications
 - Standard XY scan – channel separation, co-localization (**CONTROL SLIDE**)
 - XYZ scan – 3D – produces pretty pictures but little value in analysis (the individual sections provide a lot more information).(**ADMIN – SEED**)
 - XZ scan – particular useful for the microbiologists to measure colony depth (**CHARLTON FILE**)
 - XYt – FRAP, FRET and other live cell applications
 - XYt λ – Spectral de-convolution – separation of 2 colours if the wavelengths are known (Admin – RBC Lambda scan image)
- FRAP – Fluorescence Recovery After Photobleaching
 - Will tell us the amount of protein movement/diffusion over time (**ClaireM Image 39**)
- FRET – Fluorescence Resonance Energy Transfer
 - Will tell us if 2 proteins are likely to be interacting
 - Microscope optical resolution limit = 200nm
 - FRET only occurs within 10nm
- Live Cell Imaging
 - Need humidibox
 - Need 5% CO₂
 - Need anti-vibration table
 - Pros – get to see protein movement and protein-protein interactions as they happen
 - Con – kill the cell, lot of work involved to get excellent images
 - (**Kristine File – 260209LCI – YrdC203 RpL12**).
- Other interesting confocal stuff:

- Photoactivatable proteins.



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- Photoactivatable GFP – 405nm laser induces the expression of GFP at 488nm – useful for tracking the dynamics of molecular subpopulations within a cell.
- Kaede – Photoconversion from green to red after exposure to 405nm laser - Example: allows delineation of a single neuron in a dense culture

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References:

<http://www.dbs.nus.edu.sg/research/facilities/confocal/LSM%20510%20META%20training-DBS-070605.pdf>

<http://www.olympusmicro.com/primer/techniques/confocal/index.html>

<http://www.microscopyu.com/articles>